Pharmacological Prevention of Neonatal Opioid Withdrawal in a Pregnant Guinea Pig Model

Alireza Safa [1†], Allison R. Lau [2†], Sydney Aten [1†], Karl Schilling [3], Karen L. Bales [2], Victoria A. Miller [1], Julie Fitzgerald [1], Min Chen [4], Kasey Hill [4], Kyle Dzwigalski [4], Karl Obrietan [1], Mitch A. Phelps [4], Wolfgang Sadee [5,6]* and John Oberdick [1]*

[1]Department of Neuroscience, The Ohio State University Wexner Medical Center, Columbus, OH, United States, [2]Department of Psychology, California National Primate Research Center, Animal Behavior Graduate Group, University of California, Davis, CA, United States, [3]Anatomisches Institut, Anatomie und Zellbiologie, Rheinische Friedrich-Wilhelms-Universität, Bonn, Germany, [4]Division of Pharmaceutics and Pharmaceutical Chemistry, College of Pharmacy, The Ohio State University, Columbus, OH, United States, [5]Department of Cancer Biology and Genetics, Ohio State University Wexner Medical Center, Columbus, OH, United States, [6]Aether Therapeutics Inc., Austin, TX, United States

Newborns exposed to prenatal opioids often experience intense postnatal withdrawal after cessation of the opioid, called neonatal opioid withdrawal syndrome (NOWS), with limited pre- and postnatal therapeutic options available. In a prior study in pregnant mice we demonstrated that the peripherally selective opioid antagonist, 6β-naltrexol (6BN), is a promising drug candidate for preventive prenatal treatment of NOWS, and a therapeutic mechanism was proposed based on preferential delivery of 6BN to fetal brain with relative exclusion from maternal brain. Here, we have developed methadone (MTD) treated pregnant guinea pigs as a physiologically more suitable model, enabling detection of robust spontaneous neonatal withdrawal. Prenatal MTD significantly aggravates two classic maternal separation stress behaviors in newborn guinea pigs: calling (vocalizing) and searching (locomotion) - natural attachment behaviors thought to be controlled by the endogenous opioid system. In addition, prenatal MTD significantly increases the levels of plasma cortisol in newborns, showing that cessation of MTD at birth engages the hypothalamic-pituitary-adrenal (HPA) axis. We find that co-administration of 6BN with MTD prevents these withdrawal symptoms in newborn pups with extreme potency (ID50 ~0.02 mg/kg), at doses unlikely to induce maternal or fetal withdrawal or to interfere with opioid antinociception based on many prior studies in rodents and non-human primates. Furthermore, we demonstrate a similarly high potency of 6BN in preventing opioid withdrawal in adult guinea pigs (ID50 = 0.01 mg/kg). This high potency appears to run counter to our pharmacokinetic studies showing slow 6BN transit of both the placenta and maternal blood brain barrier in guinea pigs, and calls into question the preferential delivery mechanism. Rather, it suggests a novel receptor mechanism to account for the selectively high potency of 6BN to suppress opioid dependence at all developmental stages, even in adults, as compared to its well-established low potency as a classical opioid antagonist. In conclusion, 6BN is an attractive compound for development of a preventive therapy for NOWS.

Keywords: opioid, neonatal, withdrawal, guinea pig, preventive, therapeutic, hypothalamic-pituitary-adrenal axis, cortisol

INTRODUCTION

Extended prenatal exposure to an opioid, with cessation at birth, may result in neonatal opioid withdrawal syndrome (NOWS) (Yeoh et al., 2019). NOWS babies are often born premature and underweight, are typically extremely irritable with inconsolable high-pitched crying, with uncontrolled and jittery limb movements, disrupted sleep, and other complications. These problems result in extended ICU stay times, accounting for most of the financial costs of NOWS (Patrick et al., 2012; Patrick et al., 2015a). In addition, there are significant later-life effects, including motor and cognitive delay, with untold long-term costs (Patrick et al., 2015b; Nygaard et al., 2016). Opioids continue to be the best relief for chronic or severe pain, and as many as 28% of pregnant women are reported to have filled an opioid prescription for pain or opioid use disorder (OUD) management (Yeoh et al., 2019). Buprenorphine, a favored agonist for maintenance therapy, results in less severe neonatal withdrawal than methadone (MTD), but both agonists have a similar rate of NOWS after birth (~50%), and both require significant ICU stay times (Jones et al., 2010). Here, we evaluate a prenatal preventive strategy that, in principle, should reduce long ICU stay-times, the need for postnatal morphine treatment of NOWS babies, and the negative developmental consequences of prenatal opioids. Currently only palliative therapies are available to those babies that develop NOWS, and none would prevent developmental delay.

Peripherally selective opioid receptor antagonists, such as naloxegol, have been developed to prevent the peripheral effects of opioids such as constipation, without affecting central control of pain by an opioid (Chey et al., 2014). The relative exclusion of these compounds from the central nervous system is thought to be due to efflux transporters at the mature blood brain barrier (BBB), such as P-gp (Xie et al., 1999). In a previous study we demonstrated the preferential delivery of another peripherally selective opioid receptor antagonist, 6β-naltrexol (6BN), to the fetal brain in pregnant mice (Oberdick et al., 2016). While partially excluded from the maternal brain, 6BN rapidly transits the placenta and enters the fetal brain reaching levels 6-fold higher than in maternal brain. The developmentally regulated expression of transporters such as P-gp at the BBB may account for the observation that 6BN readily entered the fetal brain and brains of pre-weaning juvenile mice until at least postnatal day (PD) 15 (Oberdick et al., 2016). Inducing morphine dependence in juvenile mice, we demonstrated that co-administration of 6BN at extremely low doses prevents subsequent withdrawal behavior with a 50% inhibitory dose (ID50) of 0.02–0.04 mg/kg–500-fold lower than the dose of morphine used to induce dependence in the study, and 50–100-fold lower than the published ID50 of 6BN to block opiate antinociception or to induce withdrawal in opioid dependent adult animals, including in mice and monkeys (Wang et al., 2001; Porter et al., 2002; Wang et al., 2004; Raehal et al., 2005; Ko et al., 2006; Yancey-Wrona et al., 2009). Here, we test the hypothesis that 6BN can prevent fetal dependence and subsequent neonatal withdrawal when co-administered at extremely low doses with an opioid agonist–at 6BN doses too low to interfere with maternal pain or addiction management by an opioid.

Facile 6BN access to the immature mouse brain (Oberdick et al., 2016) should account for no more than 5–10-fold higher potency in blocking opioid antinociception compared to adults with an intact BBB. Therefore, it is likely that factors other than the BBB are at play, and alternative mechanisms at the opioid receptor likely play a role in 6BN's extremely high potency to prevent dependence in mouse juveniles as compared to its 50–100-fold lower potency in blocking other agonist actions in adults. We address this question here by also testing the ability of 6BN to prevent dependence and withdrawal when co-delivered with MTD to induce dependence in adult guinea pigs. Although our previous study in mice was focused on morphine as an agonist, we switched to MTD here because it is commonly used to treat OUD during pregnancy, as described above (Jones et al., 2010). This population of women would be potentially accessible for testing a preventive therapeutic for NOWS. MTD also has a much longer half-life than morphine, and could thus serve as a more stringent test of 6BN efficacy.

There are some limitations of laboratory mice and rats as models for NOWS. Mice show no reported behavioral effects of prenatal opioids at birth, but rat pups display spontaneous and naloxone-induced increased movements (Richardson et al., 2006; Griffin et al., 2019; Robinson and Wallace, 2001). The latter behaviors are strictly count-based binary measures (yes-no) of limb, body and head moves. The subtlety of these behaviors is presumably due to the relative underdevelopment of the brain of these species at birth, which are the neurodevelopmental equivalent of mid to late-second trimester human fetuses (Workman et al., 2013, and http://translatingtime.org/translate). However, more robust withdrawal behaviors in the locomotor and ultrasonic vocalization domains can be observed by PD7-10 in mouse and rat pups using postnatal exposures to an opioid (Barr and Wang, 1992; Oberdick et al., 2016). Mice and rats at this age are the neurodevelopmental equivalent of human newborns and may be useful for modeling some aspects of NOWS. But they lack the temporal continuity of opioid cessation at birth, as occurs in NOWS.

In order to develop a more human-relevant model we have been studying guinea pigs, a precocial species neurodevelopmentally more akin at birth to a human infant (Workman et al., 2013, and http://translatingtime.org/translate). These studies suggest that cessation of MTD at birth after extended prenatal exposure causes an increased "craving" for the opioidergic effects provided by infant-parent contact, resulting in NOWS, and likely engaging stress mechanisms mediated by the hypothalamic-pituitary-adrenal (HPA) axis. Finally, we show for the first time that 6BN can prevent spontaneous withdrawal behaviors after birth when co-administered with a prenatal opioid, and it can do so with high potency. Moreover, the high potency does not appear to require preferential delivery of 6BN to the fetal CNS. (This manuscript has been released as a pre-print (Safa et al., 2020)).

MATERIALS AND METHODS

Animals

Hartley guinea pigs (Cavea porcellus) were purchased from Charles River. All adult non-pregnant animals were 2–4 months old at the time of testing. Pregnant sows for PK studies were purchased in three cohorts of seven animals each. Pregnant sows for neonatal behavior studies were purchased in six cohorts of 8–10 animals per cohort. All pregnant animals were shipped at early gestation (~GD30) to reduce pregnancy-related toxemia, which is a significant concern for guinea pigs. The standard range of pup birthweights for Hartley guinea pigs is 50–100 g (https://www.criver.com/sites/default/files/resources/rm_rm_n_techbul_spring_99.pdf). However, for in-house reared guinea pigs, birthweights below 85 g are sometimes considered low birthweight, and "normal" birthweight is considered those on the high end of the normal range, >90 g (Sarr et al., 2016). Thus, the relatively lower mean birthweight of 75.5 ± 3.1 g for saline control animals in the current study may reflect vendor-related substrain differences, but we cannot rule out an impact of shipping-related stress. Pregnant sows were typically second or third pregnancies, and 6–8 month old. These were "multi untimed pregnant animals" with a 3 days window of variation for day of conception (gestation time of guinea pigs is ~65 days). Animals were pair-housed whenever possible in solid bottom open caging (Allentown, Allentown, NJ, cage pans measured 29 × 21 × 10 inches) with Sani Chip bedding (Teklad 7090, Envigo). Animals were fed a chow diet ad libitum (Teklad Guinea Pig Diet 2040, Envigo) that was supplemented with hay and a rotation of fresh produce provided daily. Reverse osmosis purified water was provided via an automated rack watering system, and a 12/12 h light/dark cycle was maintained for the duration of the experiment.

All procedures were approved by The Ohio State University Institutional Animal Care and Use Committee and are in compliance with guidelines established by the National Institutes of Health published in Guide for the Care and Use of Laboratory Animals (http://oacu.od.nih.gov/regs/guide/guide.pdf).

Drugs

6β-naltrexol (6BN) (https://pubchem.ncbi.nlm.nih.gov/compound/5486554), naltrexone (https://pubchem.ncbi.nlm.nih.gov/compound/5360515) and D,L-methadone (MTD) (https://pubchem.ncbi.nlm.nih.gov/compound/14184) were provided by the Drug Supply Program of the National Institute for Drug Addiction (NIDA) as previously reported (Oberdick et al., 2016). Drugs were dissolved in saline at concentrations between 10 and 20 mg/ml, and all dilutions were made in saline.

Pharmacokinetic (PK) Analysis

6BN and Naltrexone Dosing and Tissue Collection

For the bioavailability study young adult (2 months old) animals with jugular vein catheters were purchased from Charles River for plasma multi-sampling and IV dosing. Half of the animals were used for IV, half for oral delivery. For oral delivery animals were briefly anesthetized with isoflurane, held vertically, and drug (in saline +1% sucrose) was delivered with a feeding needle, typically in 1.5–3 ml. This procedure induced significant salivation and/or regurgitation, which contributed to dosing variability. For the PK studies in brain and plasma most of the reported data used oral delivery. However, as indicated in the Results other animals were injected subcutaneously in the dorsolateral region around the shoulders of the forelimbs. Injection volume typically did not exceed 750 ul in adults. Embryos and maternal tissues were collected for the plasma and brain PK study. After drug injection and variable survival times pregnant animals were euthanized by CO_2 followed by heart puncture (for maternal blood collection, ~0.5–1 ml) and decapitation. Fetuses were collected onto large Petri dish lids kept on wet ice, and blood was collected in a 1 ml syringe (no needle) from pooling of blood around the neck region after severing of both jugular veins (with animal on its back). Then brain and liver were collected and frozen on dry-ice while plasma was prepared in a microfuge at room temp (typically from 200–400 ul of fetal blood). All samples were stored at −70°C until processing. In the mass spectrometry lab tissues were thawed, resected, weighed, and quickly frozen again on dry ice in individual microcentrifuge tubes. Samples were later thawed, processed and analyzed via liquid chromatography-tandem mass spectrometry (LC-MS/MS) to quantify levels of drug.

Bioanalytical Assay for 6β-Naltrexol and Naltrexone

Guinea pig plasma and brain tissue were assayed according to Oberdick et al., 2016 (PMID: 27189967) with the following changes going from mouse to guinea pig plasma and tissues. Guinea pig brain and liver tissues were homogenized in 1X PBS pH 7.4 at concentrations of 66.7 and 33.3 mg/ml, respectively. Forty-5 μL of plasma or tissue homogenate were spiked with 5 μL of 3,000 ng/ml [^{2}H-7]-naltrexone or a combination of [^{2}H-7]-naltrexone and 6β-naltrexol-d3. Samples were then extracted with acetonitrile:methanol (3: 1, v/v). The post centrifugation supernatant was transferred to a new 2 ml 96-well plate then dried with a stream of nitrogen before reconstituting with 120 μL of 0.1% formic acid. The reconstituted samples were transferred to a 96-well autosampler plate for a 5 μL injection onto a Thermo Accucore Vanquish C18 + column (1.5 μm, 100 × 2.1 mm). The chromatographic system was a Thermo Scientific Vanquish Horizon UHPLC. The analytes were separated using a 3.5 min gradient elution program with 0.1% formic acid (MPA) and acetonitrile:methanol (3:1, v/v) with 0.1% formic acid (MPB) as the mobile phases. The gradient began with 10% MPB for 0.4 min then increased to 90% MPB over 1 min. Then 90% MPB was held for 0.6 min before decreasing back to 10% MPB to equilibrate the column. The flow rate for the gradient was 0.4 ml/min and the column was maintained at 40°C. The samples were analyzed on either a Thermo Scientific TSQ Quantiva or a TSQ Altis equipped with an electrospray ionization source in positive polarity. The mass transitions (m/z) monitored were 342.18 > 324.11 for naltrexone, 344.21 > 326.11 for 6β-naltrexol, 349.24 > 331.11 for [^{2}H-7]-naltrexone, and 347.36 > 329.25 for 6β-naltrexol-d3. Cross-validation included linearity, within-day, and between-day accuracy and precision. Selectivity was assessed by analyzing six lots of guinea pig plasma without and with spiking at the lower limit of quantitation (LLOQ)

levels. Recovery, matrix effects and stability were also assessed in matrix and solvent for both analytes at low and high QC concentrations. Stability was assessed on benchtop (22°C), three freeze thaw cycles from −80 to 22°C, autosampler stability for 24 h at 4°C, and long-term stability at −80°C for 30 days. The linear range, including LLOQ for 6BN and naltrexone were as follows: Plasma, x-y ng/mL 6BN, x-y ng/mL naltrexone; Liver, x-y ng/mL 6BN, x-y ng/mL naltrexone; Brain, x-y ng/mL 6BN, x-y ng/mL naltrexone.

Pharmacokinetic Data Analysis

For the bioavailability study, non-pregnant adult guinea pigs were dosed with 6BN via intravenous bolus route at 10 mg/kg and orally at 40 mg/kg. Plasma samples were collected at 0, 15, 30, 60, 120, 240 and 480 min post dose. Individual 6BN pharmacokinetic parameters were estimated from individual plasma concentration-time profiles using non-compartmental analysis (NCA) with Phoenix WinNonLin (version 8.2.0, Certara, Princeton, NJ). The terminal linear phase was identified automatically in WinNonlin using linear least squares regression to estimate the terminal elimination rate constant (λz). Area under curve (AUC) was determined using linear trapezoidal linear interpolation method. Bioavailability was calculated as the dose-normalized ratio of AUCs determined from oral and intravenous routes of administration. For the brain distribution analysis, the concentration-time profile for both brain and plasma were anlyzed with NCA methods using Phoenix WinNonLin and PK parameters were calculated in a similar fashion. Pregnant guinea pigs were dosed with 6BN orally at 40 mg/kg, then both fetal and maternal brain and plasma samples were collected at 15, 30, 60, 120, 240 and 480 min post dose (one time point per animal).

Protein Binding by Equilibrium Dialysis
Data Were Collected and Analyzed by Covance

Instrument. The high throughput dialysis apparatus, model HTD96b, was used (HTDialysis LLC, Gales Ferry, Connecticut). Prior to assembly, dialysis membrane strips (molecular weight cutoff of 6–8 kDa) were hydrated according to manufacturer's recommendations. The Teflon bars were assembled according to manufacturer's instructions, with dialysis membrane strips laid between bars creating two compartments per well. The assembled unit was locked in place in the steel base plate. Samples were immediately added to each compartment to prevent dehydration of the membranes. Equilibrium Dialysis Procedure. Fortified matrix (plasma) was added to the donor side, DPBS was added to the receiver side of the HTD wells, and the plate was sealed. Samples were incubated at 37°C and rotated at 300 rpm for the designated time. After incubation, the seal was removed and a sample from each plasma and dialysate chamber was analyzed by LC-MS/MS. All protein binding determinations were performed in quadruplicate. Time to Equilibrium Determination. Dialysis was performed according to the equilibrium dialysis procedure for 3, 5, 6, 7, and 8 h to determine the minimum time to achieve equilibrium. This experiment was conducted at 50 ng/ml of 6-beta naltrexol in human plasma. Equilibrium was

obtained when the percentage of 6-beta naltrexol bound to the proteins in the plasma remained constant over time. Concentration Dependence. The protein binding in mouse, rat, guinea pig, dog, and human plasma was determined at concentrations of 0.5, 1.5, 5, 15, and 50 ng/ml of 6-beta naltrexol. The dialysis time for the test article was 8 h as determined in the time to equilibrium experiment. Sample Analysis. Samples were processed prior to LC-MS/MS analysis as follows. The donor side samples (plasma) were diluted with DPBS and the receiver side samples (DPBS) were diluted with blank control plasma at the appropriate volumes to provide a common analytical mixed matrix of 90% DPBS and 10% plasma (90:10 DPBS:plasma, v:v) in 100 μL total volume. Samples were prepared for analysis and analyzed for 6-beta naltrexol using a quantitative LC MS/MS method developed at Covance and modified as appropriate for study optimization.

MTD and 6BN Dose-Response Studies in Newborn and Adult Guinea Pigs
Experimental Design for Newborn Studies

Overview: In this study 10 mg/kg was identified as the maximum tolerable dose of MTD for both sow and fetus survival. Thus, we determined the prenatal MTD dose response for neonatal withdrawal in the range from 0–10 mg/kg. Then we selected two MTD doses near the mid-point of the dose-response curve for testing whether 6BN could prevent the neonatal effects of MTD.

Dosing groups and animal numbers: All behavioral studies were performed between 12–4 PM to ensure similar circadian schedules. Generally, tests were performed in cohorts of 20–30 pups from 8–10 sows at the same gestational stage (with a 3 days window of variation; see Animals section). With sow numbers thus restricted pups were born over the span of 1 week, which allowed us to maintain the 4 h testing window each day. Each cohort was designed with multiple dosing groups, always including either a saline control group and/or a MTD only (no 6BN) control group. Pregnant sows were pair-housed, and both animals in a pair received the same treatment.

23 pregnant animals were used for the MTD dose response study, and 37 pregnant animals were used for the 6BN dose response study. No more than three and an average of two pups per litter were used for all withdrawal analysis to minimize litter effects (for all pregnant animals with all treatments the average litter size was 4.7 ± 1.7 (±SD)). Pups for the MTD dose study came from at least two litters per dose group with an average of 4 litters per group. Groups for the MTD study were saline controls ("0" MTD), and 2, 5, 7, and 10 mg/kg MTD. For the analysis of the effects of 6BN, data from animals treated with 5 mg/kg MTD (123 pups; five to seven litters per 6BN dose group) and 7 mg/kg MTD (30 pups, 2 litters per dose of 6BN) were pooled. This approach seems justified since despite the general MTD-dose dependence of behavior that may be discerned if the whole dose range of 0–10 mg/kg is considered the behavior of animals treated with

five or 7 mg/kg of MTD was not detectably different, and the same is true of animals treated with 0 or 2 mg/kg MTD. We also note that no statistical interactions between MTD and 6BN could be detected in this combined group. Finally, we note that analysis of the effects of 6BN in the animals treated with 5 mg/kg prenatal MTD yielded very similar results to what was observed for the combined 5 and 7 mg/kg MTD animals, with the obvious differences as one may expect due to a smaller group size.

Injection schedule: Pregnant animals were received in the vivarium on ~ GD33, acclimated for 17 days, and daily single injections of saline, MTD, or MTD with 6BN at variable doses were initiated at GD50 for an average of 15 ± 3 (±SD) injections before birth at ~ GD65. Pups were marked with an indelible marker on the inside of the ear 24 h after birth. Pups were behaviorally tested (locomotion and vocalization) at 48 h after birth (see below).

Adult Withdrawal Testing

15 females (non-pregnant) and 11 males were used for this study, 2–4 months of age. As for the newborns they were tested in the open-field (see below). Vocalization testing was not performed.

Behavioral Tests

Open-field locomotor testing: Open field tests were performed on pups 48 ± 12 h after birth in order to examine their locomotor behavior before and after naloxone administration. One hour before testing the entire home-cage was transported from the housing room into an enclosed holding vestibule with a door to a separate behavior room. Two cameras (Panasonic model SDR-H80 and Sony Handycam model DCR-SR45) were used to record the animals. One camera (Panasonic) was mounted directly over the open field arena on a stand at a fixed height (104 cm) for all studies. The zoom on the camera was adjusted so that the top and bottom borders of the square arena filled the entire height dimension of the camera's rectangular field of view. This was the main camera used for video and acoustic analysis. The other camera was placed on the ground adjacent to the arena in order to examine detailed facial expressions and behaviors. The arena was 40 by 40 cm in size, and clean wood chip bedding was placed on the floor of the arena (same type as for housing). Animals were removed from the home-cage in the vestibule, brought into the testing room, and weighed immediately before being placed in the open field arena. Once in the arena they were video-recorded for 10 min. Then they were removed from the arena, injected with naloxone (s.c.) at a dose of 20 mg/kg, and immediately placed back in the arena and video-recorded for 30 min. Note that no more than three pups per litter were run in the open field test, with an average of two pups per litter. Also, at the start of each video an index card with the date and animal ID was placed briefly in the field of view of the camera and recorded. All videos are archived on external hard-drives and are available to collaborators on an online file sharing service.

Locomotor testing of adults was performed in precisely the same manner using a 10 min open-field test prior to naloxone, followed by injection with naloxone, and a 30 min test in the same arena.

Video analysis: Each video was imported into the ANY-maze software (version 6.06), labeled by date and animal ID, and data extracted by an observer blinded to animal treatment. In each video the area of the open field chamber was manually defined using a drawing tool, and a distance of known length within this area was used for standardization. The distance traveled by each animal was calculated based on the displacement of the center point of their bodies (the white coat color of animals was easily detected against the darker wood shavings on the floor of the apparatus). Freezing detection was set up using the default values in the software: a minimum freeze duration of 1,000 milliseconds, a "freezing on" threshold of 30, and a "freezing off" threshold of 40. Once all the parameters were entered in the software, the videos were run for analysis. Data from each animal was then assigned to its proper dosage group by another researcher for statistical analysis.

Vocalization analysis: WAV audio files were extracted from the same videos that were used for automated video analysis, then analyzed using Raven Pro 1.6 Sound Analysis Software (Center for Conservation Bioacoustics, Cornell Lab of Ornithology, Ithaca, NY) by one researcher who was blinded to the dosage groups. Spectrograms with a 512-point (11.6 m s) Hann window (3 dB bandwidth = 124 Hz), with 75% overlap, and a 1,024-point discrete Fourier transform, yielding time and frequency measurement precision of 2.9 m s and 43.1 Hz were generated. Sounds files were not down sampled.

The features estimated were vocalization count and signal-to-noize ratio (SNR). SNR is the amplitude of the vocalization signal above the background noise. In other words, SNR measured how loud the guinea pigs were, controlling for background noise. In order to count the number of vocalizations emitted by each guinea pig pup in the 10-min observation in an objective manner, we used the Band Limited Energy Detector to select each vocalization. The Band Limited Energy Detector was tailored to detect the vocalizations of infant guinea pigs. The final detector was validated visually and audibly by one observer (ARL). The target signal parameters for our detector were minimum frequency: 390 Hz, maximum frequency: 920 Hz, minimum duration: 0.032 s, maximum duration: 0.16 s, and minimum separation: 0.032 s. We set our SNR ratio parameters to 45% minimum occupancy with an SNR threshold of 6.0 dB above. In order to estimate SNR of the sound file, one 10-min selection was generated using the Raven Pro selection tables. SNR was automatically estimated by Raven Pro 1.6.

Statistical Procedures for Dose-Response Studies

Summary data are presented as the means ± S.E.M. unless otherwise indicated. Two-sided t-tests allowing for variance heterogeneity were used to contrast samples if only two groups were to be compared. To compare multiple groups, and to screen for possible dose-response effects, we used the step-down Tukey trend test adjusted for multiplicity ((Tukey et al., 1985); referred to as Tukey trend test; also known as the Tukey-Ciminera-Heyse trend test). Essentially, this comprises a one-way ANOVA with post-hoc Dunnett tests and the simultaneous fitting of linear, logarithmic and ordinal regressions to the data, while controlling for the multiple testing involved. Again, testing

was done such as to account for variance heterogeneity between groups. In the newborn withdrawal studies sex was not considered as a factor due to the difficulty of accurately evaluating sex. In the adult withdrawal study sex was considered as a factor and showed no significant effect or interactions. Dose-response curves were fitted using an emax model (Bretz et al., 2005). These procedures were implemented in R (Schaarschmidt and Ritz, 2020) using the packages car (Fox and Weisberg, 2019), multcomp (Hothorn et al., 2008), tukeytrend (Schaarschmidt and Ritz, 2020), sandwich (Zeileis, 2004), and DoseFinding (Pinheiro et al., 2014).

The different behaviors tested capture distinct, but correlated, measures of a multifaceted response. To combine them into one common measure, we z-transformed (mean = 0; 1SD = 1) the values of each of the four behavioral measures obtained for all animals (irrespective of their treatment). This puts the values of all behavioral measures on the same scale without distorting their distribution. A composite score ("normalized behavior score") was then calculated for each animal by adding up the transformed values of the distinct behavioral measures. As all measures except freezing time decreased with increasing doses of 6BN, ranks for freezing time were multiplied by −1 (i.e., inverted) before transformation. This approach was motivated by the goal to obtain a more comprehensive and hopefully more robust characterization of behavior than might be achieved by a single endpoint. It was also motivated by the clinical scores constructed from multiple measurements, and the success of multiple-endpoint analyses in clinical studies (e.g., (Offen et al., 2007; Ristl et al., 2019; Follmann et al., 2020), to cite but a few).

We add that if data were put on comparable scales by ranking instead of z-transformation, results fully consistent with those reported above were obtained.

Analysis of Birthweight and Maternal Weight Gain

All data for this analysis were extracted from all pups from all litters described above in the MTD and 6BN dose-response studies. Pregnant sows were weighed at the start of dosing, then every 3–4 days, and then every day as they neared term. Pups were weighed at 24 and 48 h after birth. The birthweight data reported here are at 24 h.

Blood Collection and Cortisol Measurement

All samples were collected 48 ± 12 h after birth. Non-survival trunk blood was collected from pups after 1 h of maternal separation stress. For the 1 h separation we removed pups from the home-cage and placed them in a holding cage (plastic mouse cage with lid and air filter) in a different room. Pups were then sacrificed by CO_2 asphyxiation followed by rapid decapitation in the vivarium necropsy room. Trunk blood was collected (from the site of decapitation) and put into 1.5 ml Eppendorf tubes coated with EDTA (pH 8.0; Invitrogen Corporation Cat No. 15575–038) to prevent coagulation. Tubes were then centrifuged for 10 min at 2,500 rpm. Plasma was placed into a separate tube and stored at -80.

Plasma cortisol was assayed by enzyme immunoassay (antibody R4866, produced by UC-Davis Endocrinology Laboratory (see Bales et al., 2005; Bales et al., 2006) at a 1: 1,000 dilution. Samples were assayed in duplicate and high and low pools were run with each assay. As all samples were run in one assay, there are no inter-assay cv's to report. The low controls had an intra-assay cv of 3.61%, while the high controls had an intra-assay cv of 3.58%. The assay was chemically validated for guinea pigs by assessing parallelism and quantitative recovery.

RESULTS

Pharmacokinetics of 6β-Naltrexol in Fetal and Maternal Guinea Pig

We first determined the oral bioavailability of 6BN and other PK parameters in non-pregnant adult guinea pigs (**Table 1**; **Figure 1A**). 6BN is orally bioavailable with F = 29%, and t1/2 is 1.1 h for IV delivery, 2.1 h by oral delivery. For comparison methadone has a bioavailability range of 36–100% in humans with great interindividual variation, and t1/2 of 12 h in guinea pigs and 15–207 h in humans (Pak and Ecobichon, 1978; Bart et al., 2014; Eap et al., 2002; see also https://www.drugbank.ca/drugs/DB00333)–the t1/2 of 6BN is 12 h in humans (Yancey-Wrona et al., 2011). In addition, plasma protein binding of 6BN in all species tested is low over a 6BN concentration range from 0. 5–50 nM (see **Table 2**; mean unbound fraction±SD = 91 ± 6% in guinea pig, 95 ± 5% in mouse, 92 ± 11% in dog, and 90 ± 5% in human). For comparison, the published plasma protein binding of methadone is much higher (mean unbound fraction 11–14%; (Eap et al., 2002)).

We next examined the time course of tissue distribution of 6BN using oral delivery (40 mg/kg; **Figure 1B** and **Table 3**). By 1 h after dosing 6BN reaches a peak in maternal plasma that is ~4-fold higher than in fetal plasma, and fetal brain levels are ~1.5-fold higher than maternal brain levels at 1–2 h (see AUC_{0-4} in **Table 3**). In addition, fetal plasma and brain levels are near equivalence at peak in the first 120 min; this result was replicated at a 10-fold lower oral dose of 4 mg/kg 6BN. Using combined data at the two 6BN doses the fetal brain/plasma ratio is 1.04, while the maternal brain/plasma ratio is 0.220 ($p < 0.001$ for comparison of fetal vs maternal brain/plasma ratio, by t-test). In addition, the absolute level of 6BN in fetal brain in guinea pigs is ~6-fold lower than in mouse fetal brain under conditions with roughly equal maternal plasma 6BN levels; maternal brain levels are roughly the same across the two species (compare data in **Figure 1B** to that in (Oberdick et al., 2016)). These results demonstrate slow placental transfer of 6BN in guinea pigs, but with rapid fetal brain entry, and relative exclusion from maternal brain. Thus, compared to mice, preferential delivery of 6BN to fetal brain is lower in guinea pigs than in mice due to slow placental transfer (i.e., fetal/maternal AUC ratio of brain levels is 1.5 in guinea pig vs six in mouse; mouse data reported previously in (Oberdick et al., 2016)). Due to high variability with oral dosing in pregnant guinea pigs (see Methods) we repeated some of this analysis using subcutaneous delivery with both single injection and multi-injection paradigms. Over a dose

TABLE 1 | Plasma PK analysis and bioavailability of 6BN.

Parameters	Unit	IV (10 mg/kg)	PO (40 mg/kg)
		n = 3	n = 4
T1/2	hr	1.12 (0.75,1.43)	2.06 (1.17,2.68)
C0	ng/ml	3,280 ± 3,300	
Cmax	ng/ml		647 ± 279
Tmax	hr		0.25 (0.25,0.5)
VSS	L/kg	7.27 ± 4.29	
CL	ml/min/kg	132 ± 56	
AUClast	hr*ng/ml	1,450 ± 861	1,620 ± 540
AUCinf	hr*ng/ml	1,490 ± 830	1710 ± 513
Bioavailability[a]			28.7

[a]Bioavailability was calculated with AUCinf and nominal doses.
Values are geometric mean (geometric SD) except for t1/2 and T_{max} values are median (min, max).

range from 0.5–10 mg/kg, regardless of the dosing schedule and route, we found slow placental transfer of 6BN in pregnant guinea pigs, similar to that in **Figure 1B** (data not shown). In addition, while naltrexone showed similar maternal plasma levels as 6BN at the same dose and time after administration, we observed ~10-fold higher levels of naltrexone compared to 6BN in both fetal and maternal brain (**Supplementary Table S1**), indicating more rapid placental transfer and higher maternal brain levels of naltrexone compared to 6BN. These results are consistent with reported relative exclusion of 6BN from adult brain as compared to naltrexone based on pharmacokinetic studies (Wang et al., 2004; Oberdick et al., 2016), and seem to correlate qualitatively, albeit not quantitatively (as discussed in the Introduction), with the greater potency of naltrexone to block opioid analgesia and/or to induce withdrawal in opioid dependent adult animals (Wang et al., 2001; Porter et al., 2002; Wang et al., 2004; Raehal et al., 2005; Ko et al., 2006; Yancey-Wrona et al., 2009).

Prenatal Methadone Aggravates Maternal Separation Stress Behaviors in Newborn Guinea Pigs
Methadone Effect on Maternal Weight Gain and Pup Birthweight

In this study we focused our attention on MTD since it is commonly used in the clinic for opioid maintenance therapy (Jones et al., 2010), and such patients could be an accessible target population for testing a 6BN therapy. The standard analgesic dose of MTD for guinea pigs is 3–6 mg/kg (Hawk et al., 2005). However, in a previous study on respiratory effects of prenatal MTD in newborn guinea pigs, 12 mg/kg was found to be the highest dose that did not result in lethality of the pups (Nettleton et al., 2008). Therefore, as part of a similar effort to establish the maximum dose of MTD we performed a pilot study on two pregnant animals at this dose. One lost ~13% of her body weight and was lethargic and unresponsive after 6 days of dosing, and had to be euthanized. The other also lost ~13% body weight, but over a longer time, gave birth prematurely, and all pups died. We then focused on the prenatal MTD range from 2 mg/kg to 10 mg/kg, first examining maternal weight gain during pregnancy and pup birthweight. As shown in **Table 4** control
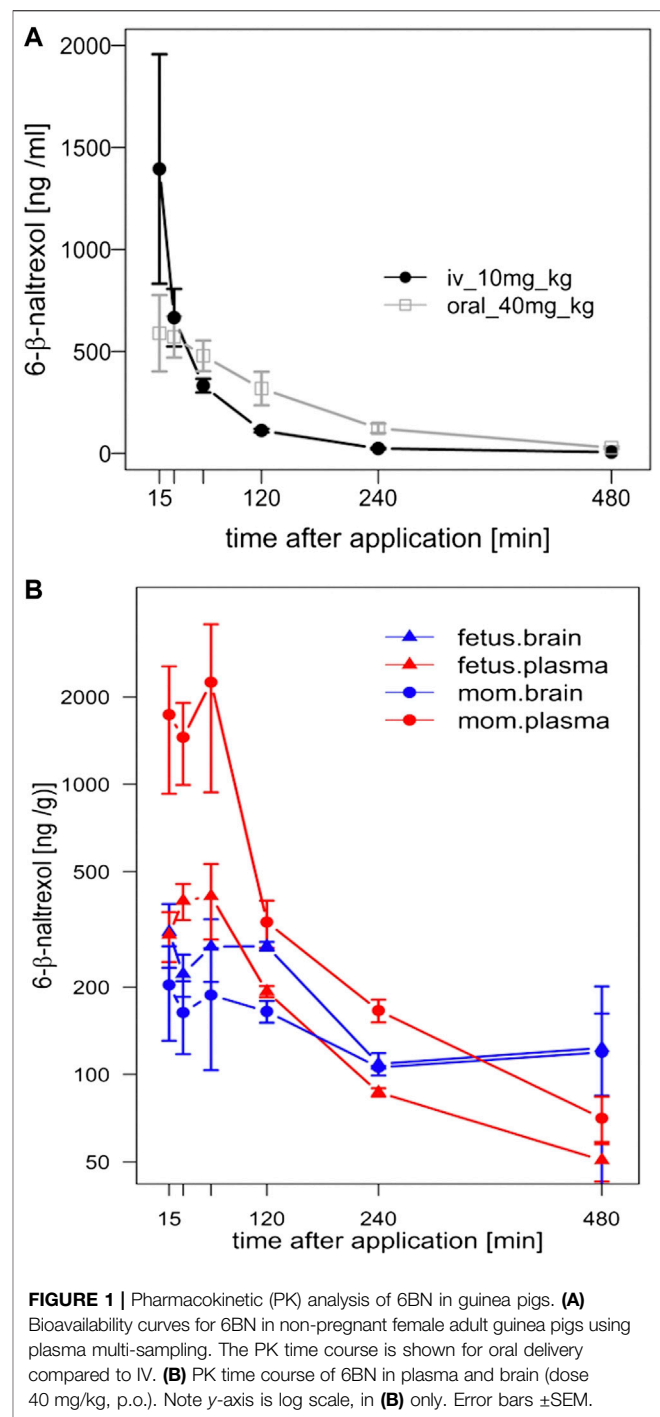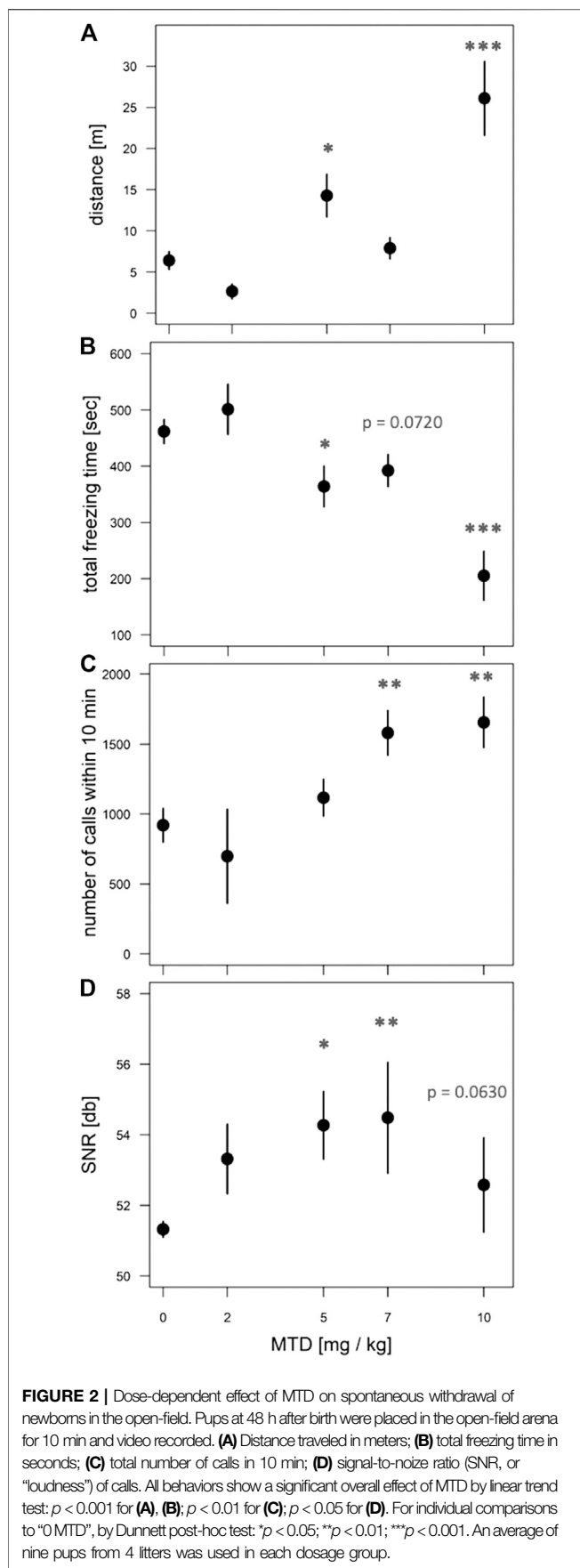

FIGURE 1 | Pharmacokinetic (PK) analysis of 6BN in guinea pigs. **(A)** Bioavailability curves for 6BN in non-pregnant female adult guinea pigs using plasma multi-sampling. The PK time course is shown for oral delivery compared to IV. **(B)** PK time course of 6BN in plasma and brain (dose 40 mg/kg, p.o.). Note y-axis is log scale, in **(B)** only. Error bars ±SEM.
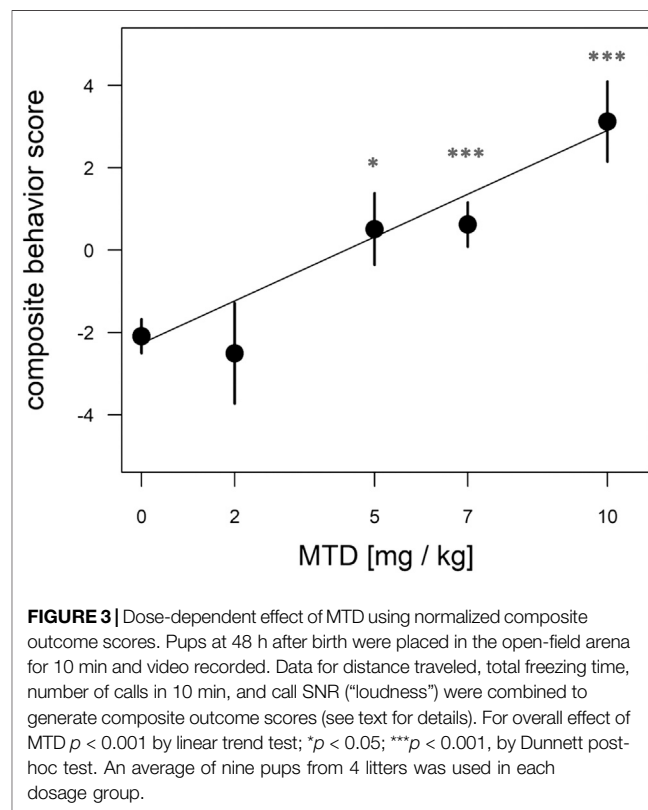
sows gained an average of 206 ± 34 g from the first saline injection to the last injection before birth (average of 15 injections). MTD resulted in a significant dose-dependent decrease in maternal weight gain relative to saline controls with a trend toward reduced weight gain at 2, 5, and 7 mg/kg, no weight gain at 10 mg/kg, and average weight loss of −175 ± 6 g at 12 mg/kg (**Table 4**; $F (5,18) =$ 55.7, $p < 0.0001$, for overall effect of MTD on maternal weight gain, by ANOVA). Also shown in **Table 4**, pup mortality

FIGURE 2 | Dose-dependent effect of MTD on spontaneous withdrawal of newborns in the open-field. Pups at 48 h after birth were placed in the open-field arena for 10 min and video recorded. (A) Distance traveled in meters; (B) total freezing time in seconds; (C) total number of calls in 10 min; (D) signal-to-noize ratio (SNR, or "loudness") of calls. All behaviors show a significant overall effect of MTD by linear trend test: $p < 0.001$ for (A), (B); $p < 0.01$ for (C); $p < 0.05$ for (D). For individual comparisons to "0 MTD", by Dunnett post-hoc test: *$p < 0.05$; **$p < 0.01$; ***$p < 0.001$. An average of nine pups from 4 litters was used in each dosage group.

increased sharply at the 10 and 12 mg/kg MTD dose, but too few animals were analyzed for meaningful statistical analysis. For pup birthweight, those exposed to 5–10 mg/kg methadone showed no difference from saline controls, but those at 2 mg/kg had a significantly *higher* birthweight than all others (n = 6 pups from 2 litters; **Table 5**). Thus, under the conditions of this study MTD had a robust effect on maternal weight gain with little effect on pup birthweight. However, low-dose MTD may lead to increased birthweight, an observation requiring further study since, while significant, there were only 2 litters.

MTD Effect on Neonatal Withdrawal-Related Behaviors

To test the effect of prenatal MTD on newborn guinea pig behavior we performed dose-response analysis, testing locomotor and vocalization behavior in an open-field arena (see Methods). The most robust effect of MTD was observed in the 10 min test (before naloxone administration). All newborns, including saline controls, displayed intense spontaneous locomotion immediately upon being placed in the arena. In addition, the pups routinely produced an audible high-pitched call (see **Supplementary Movie S1**). These are maternal separation behaviors that have been previously described, consisting of an initial active seeking and calling phase, followed by a more protracted "despair" phase (Pettijohn 1979; Hennessy et al., 2001; Hennessy 2003). The effect of MTD was examined at four doses: 2, 5, 7, and 10 mg/kg. A clear dose-dependent effect of MTD was observed in each of four separate measures: two locomotor measures (distance traveled in the arena and total time
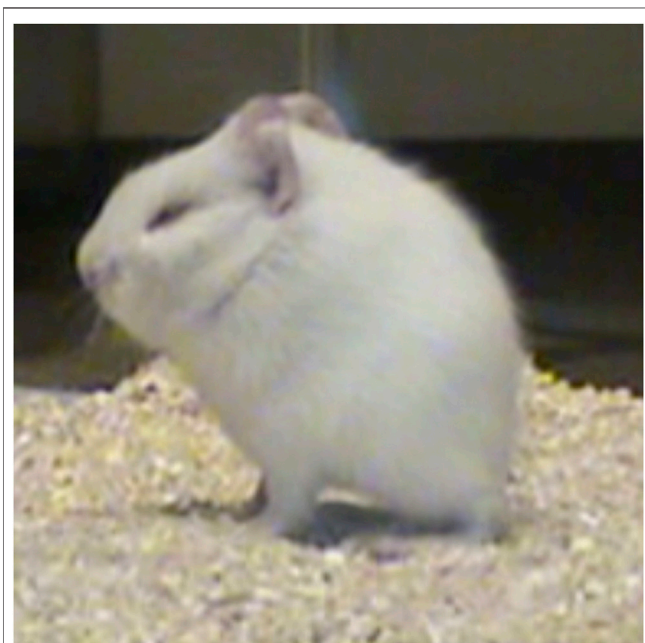

FIGURE 3 | Dose-dependent effect of MTD using normalized composite outcome scores. Pups at 48 h after birth were placed in the open-field arena for 10 min and video recorded. Data for distance traveled, total freezing time, number of calls in 10 min, and call SNR ("loudness") were combined to generate composite outcome scores (see text for details). For overall effect of MTD $p < 0.001$ by linear trend test; *$p < 0.05$; ***$p < 0.001$, by Dunnett post-hoc test. An average of nine pups from 4 litters was used in each dosage group.

FIGURE 4 | An example of immobility, piloerect posture and eye closure after naloxone induction of withdrawal in newborn guinea pigs exposed to prenatal MTD.

freezing) and two measures of vocal behavior (number of calls in 10 min and signal-to-noise ratio (SNR: defined as the amplitude of the vocalization signal above the background noise; or in other words, how "loud" the guinea pigs are, controlling for background noise) (**Figure 2**). To take better advantage of all data we also devised a composite outcome score for each animal based on z-transformation of all four

measures (see Methods). This approach was modeled after clinical studies using scores constructed from multiple measurements or endpoints (e.g., see (Teixeira-Pinto et al., 2009)). As shown in **Figure 3**, this approach not only *improved the continuity* of the dose-response ($p < 0.0001$ for the linear fit), but also it improved detection of a significant effect at all three of the highest doses (note asterisks in **Figure 3**, and compare to **Figure 2**).

In contrast, after naloxone, many pups in all groups remained nearly completely immobile during the entire period of recording (**Figure 4**). The posture of immobile animals is reminiscent of the piloerection behavior described for the late or despair phase of maternal separation stress in many studies in guinea pigs and monkeys (Hennessy et al., 2001). By contrast, other pups in some of the same groups showed classic naloxone-induced hyperlocomotion, and on rare occasions even jumping (not shown). Together the two contrasting behaviors of locomotion and immobility create a highly variable behavioral phenotype, and there is no overall significant effect of MTD in any measure after naloxone treatment (although there is a trend toward increasing locomotion distance with increasing MTD; $F_{(4,42)} = 2.38$, $p = 0.0674$). In addition, if all naloxone data from the entire study are pooled, locomotion distance was significantly decreased in the 30 min test after naloxone compared to the 10 min test before naloxone (mean ± SEM = 10.7 ± 1.0 m before naloxone vs 6.5 ± 1.2 m after naloxone, and $F_{(1,202)} = 7.06$, $p = 0.0085$), which is not the expected result for a classic naloxone induced withdrawal response.

The above naloxone effect in newborns differs from adult and juvenile mice, which show robust naloxone-induced locomotion and jumping after several days of opioid exposure (Oberdick et al., 2016; Wang et al., 2001; Wang et al., 2004; Kest et al., 2002). Here, we tested adult guinea pigs after 3 days of exposure to
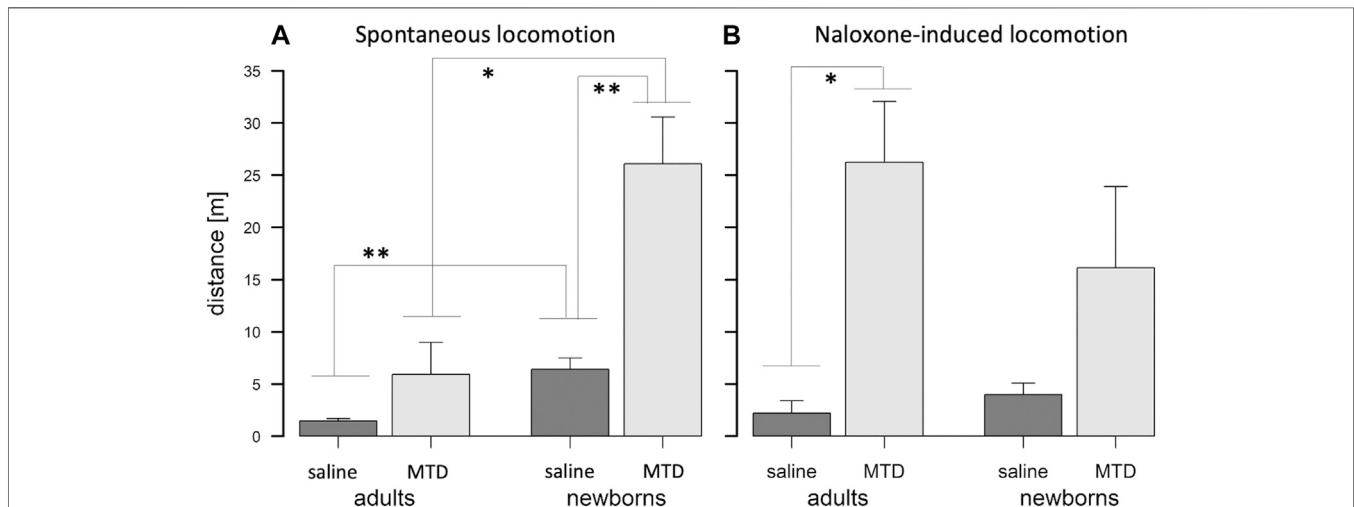

FIGURE 5 | Dose-dependent effect of 6BN using normalized composite outcome scores. Pups at 48 h after birth were placed in the open-field arena for 10 min and video recorded. **(A)** Data for distance traveled, total freezing time, number of calls in 10 min, and call SNR ("loudness") were merged to generate composite outcome scores (see text for details). **(B)** Same as **(A)**, but without "number of calls" as a factor in composite score. In **(A)**, **(B)**: $p < 0.05$ for overall effect of 6BN by linear trend test; *$p < 0.05$ compared to "0 6BN", by Dunnett post-hoc test; ᵠᵠ$p < 0.01$ for "0 6BN" compared to saline control (gray lines, mean ± SEM). An average of 16 pups from 8 litters was used in each dosage group.

TABLE 2 | 6BN mean plasma unbound fraction.

[6BN], nM	Mouse	SD	Rat	SD	Guinea pig	SD	Dog	SD	Human	SD
0.5	96.5	3.36	83.5	13.10	93.7	5.53	93.2	9.40	94.3	5.22
1.5	96.4	3.08	90.7	7.60	82.0	3.17	90.3	11.30	90.5	6.93
5	90.5	6.52	91.3	4.83	94.5	8.54	94.7	6.36	95.7	5.13
15	97.2	4.11	87.4	10.00	89.4	8.37	90.6	13.60	77.2	0.53
50	92.7	5.22	92.4	9.27	95.3	5.85	90.1	12.40	93.6	7.56
avg	94.7		89.1		91.0		91.8		90.3	

TABLE 3 | PK analysis of 6BN in fetal and maternal plasma and brain.

Parameters	Unit	Mom		Fetus	
		Plasma	Brain	Plasma	Brain
T1/2	hr	2.74	11.5	3.29	6.11
Cmax	ng/mL	2,250 (±1,310)	204 (±73)	412 (±120)	310 (±77)
Tmax	hr	1	0.25	1	0.25
AUClast	hr*ng/mL	3,810 (±1,022)	1,060 (±179)	1,190 (±96.1)	1,360 (±101)
AUCinf	hr*ng/mL	4,090	3,040	1,430	2,440
AUC0-4	hr*ng/mL	3,330	606	910	891

*Values are from same data shown in **Figure 1B**.*

TABLE 4 | Effect of methadone on maternal weight gain and litter size.

MTD dose (mg/kg)	n =	Start weight (g)	End weight (g)	Weight gain (g)	% Weight gain	AVG litter size	Total pup #	Total pup deaths
2	2	1,084 ± 40	1,221 ± 61	136 ± 21[b]	12.5 ± 1.5	3.0 + 1.0	6	0
5	8	1,195 ± 45	1,312 ± 64	118 ± 27[b]	9.7 ± 2.0	5.0 + 0.4	42	1
7	2	1,174 ± 50	1,295 ± 115	121 ± 65[b]	10.1 ± 5.1	4.0 + 1.0	8	0
10	5	1,214 ± 28	1,205 ± 11.9	−9.2 ± 24.4[a]	-0.6 ± 2.0	3.8 ± 0.7	19	7
12	2	1,335 ± 11	1,160 ± 5	−175 ± 6[a]	-13.1 ± 0.3	ND	5	5
0 (saline ctl)	5	1,188 ± 36	1,395 ± 31	206 ± 34[b]	17.7 ± 3.4	5.8 + 0.6	29	3

All values ± SEM. F(5,18) = 55.66, p < 0.0001, for overall effect of MTD on maternal weight gain. χF(3,20) = 89.66, p < 0.0001, for overall effect of MTD if data at 2, 5, and 7 mg/kg are pooled as a single group, by ANOVA.
[a]*p < 0.001 compared to saline controls, by Dunnetts posthoc test.*
[b]*p = 0.056 compared to saline controls, by Dunnetts posthoc test, if data at 2, 5, and 7 mg/kg MTD are pooled as a single group for ANOVA.*

TABLE 5 | Effect of methadone on pup birthweight.

MTD dose (mg/kg)	Number of pups (n)	Number of litters	Birthweight (g)
2	6	2	95.7 ± 7.1[a]
5	41	7	75.4 ± 1.8
7	8	2	72.4 ± 4.3
10	11	3	78.4 ± 4.0
0 (saline ctl)	28	5	75.5 ± 3.1

All values ± SEM. No significant overall effect of MTD by ANOVA: F (4,89) = 1.8671, p = 0.1232. [a]p < 0.05, for comparison to saline control, by Dunnetts posthoc test.

methadone (10 mg/kg/day). On day 4 they showed no significant increase in spontaneous locomotion relative to controls in a 10 min test in the open-field, but similar to mice, they show a robust increase in locomotion after naloxone injection; the converse of what was observed in newborns (**Figure 5**). Thus, newborn guinea pigs are unusual in not showing a significant naloxone-induced locomotor effect. This is in contrast to newborn rats exposed to a prenatal opioid, which display significant naloxone-induced increases in counts of limb and head movements (Griffin et al., 2019). In addition, the effect of prenatal MTD on spontaneous locomotion in newborn guinea pigs is an enhancement of a unique natural behavior for them, since newborn saline controls have a significant 6-fold higher level of locomotion than adult saline controls (**Figure 5A**).

In sum, these observations support that the spontaneous increase in locomotion and calling observed in neonates is due to prenatal MTD's effect on neonate-specific behaviors related to maternal separation, and not a general effect on locomotion and vocalization.

Naloxone-Induced Hypotonia

In association with the immobility behavior often observed in newborns after naloxone dosing there is an apparent sleep-related hypotonia (**Supplementary Movie S2**). The behavior always
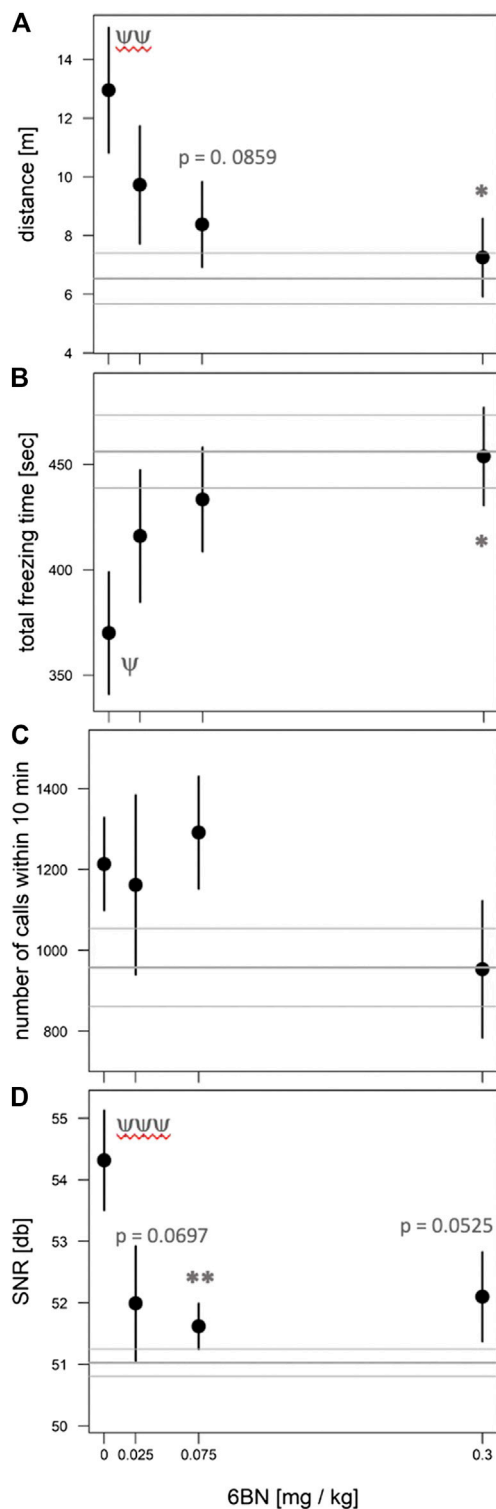
FIGURE 6 | 6BN prevents the effects of MTD on newborn withdrawal. Pups at 48 h after birth were placed in the open-field arena for 10 min and video recorded. **(A)** Distance traveled in meters; **(B)** total freezing time in seconds; **(C)** total number of calls in 10 min; **(D)** signal-to-noize ratio (SNR, or "loudness") of calls. Three behaviors show a significant overall effect of 6BN by linear trend test: *p* < 0.05 for **(A)**, **(B)** and **(D)**. For individual

(Continued)

initiates in animals that are immobilized, and in an upright and slightly hunched posture. The behavior comes in two forms: either a brief wobble and the animal rapidly rights itself (an event), or the animal collapses on its side (a "gran mal" event). After collapse the animal may stay that way for a few seconds or even a few minutes, or right itself immediately. The behavior may be associated with yawning. Efforts to quantify this behavior by counting events plus gran mals, or based on the summed duration of all events and gran mals, showed no significant effect of MTD, either when the behavior was assessed on its own or when coupled to other behaviors such as locomotion using a composite score approach. Indeed, such events are even detectable in saline control animals receiving naloxone (and in all groups receiving MTD and 6BN together; see below). They were never observed in the 10 min videos before naloxone. In total, 74 out of 91 newborns (81%) in this study treated with either prenatal saline, prenatal MTD alone or with MTD plus 6BN (see below) showed at least one event, with an average of 3.8 events per animal, after treatment with naloxone. This naloxone-induced behavior is also observed in adults exposed to MTD, but it is much less prominent than for newborns (see below). In addition, the adult data support this behavior as a contributing factor in increased variability in locomotion and other measures after naloxone.

6BN Prevents Methadone's Effect on Newborn Withdrawal Behaviors

We had reported that the partial exclusion of 6BN from the adult mouse brain was developmentally regulated and remained incomplete until 15–20 days after birth (Oberdick et al., 2016). Using PD12–17 days juvenile mice as a model for fetal drug exposure in humans, we observed that 6BN had extreme potency to prevent morphine-induced dependence and withdrawal (Oberdick et al., 2016). Also, in the same study we showed that brain AUC's of 6BN were 6-fold higher in the fetal than the maternal brain at comparable blood levels owing to the immature BBB; whether this would result, as for juveniles, in high potency of 6BN to prevent dependence could not be tested, however, since mice at birth do not show evident withdrawal after prenatal opioid exposures (Richardson et al., 2006). Our goal is to test this in guinea pigs. However, because of slow placental transfer in pregnant guinea pigs, 6BN reaches only low absolute levels in fetal brain compared to mice, and roughly equal AUC's in fetal vs. maternal brain after a single 6BN dose (see **Figure 1**, this study). Therefore, based on a preferential delivery model whereby 6BN rapidly enters fetal brain but is relatively excluded from maternal brain, we would predict decreased 6BN potency for preventing neonatal withdrawal in pregnant guinea pigs due to slow placental transfer, as compared to

FIGURE 6 | comparisons to "0 6BN", by Dunnett post-hoc test: *p* < 0.05; **p** < 0.01. For comparison of "0 6BN" to saline control (gray horizontal lines, mean ± SEM) by *t*-test: Ψ*p* < 0.05; ΨΨ*p* <0.01; ΨΨΨ*p* <0.001. An average of 16 pups from 8 litters was used in each dosage group.
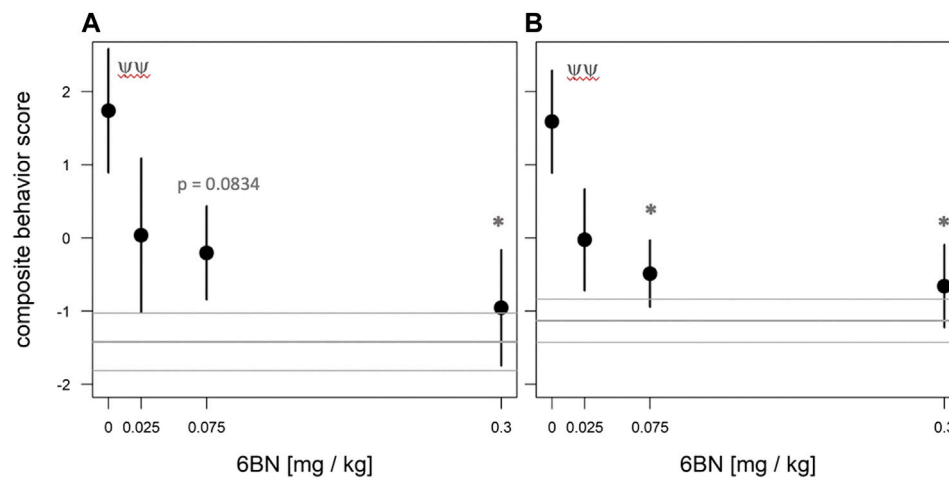
FIGURE 7 | Spontaneous vs naloxone-induced withdrawal in adult and newborn guinea pigs. **(A)** Adults treated with MTD show a trend, but no significant spontaneous withdrawal. Newborns with prenatal MTD show significant spontaneous withdrawal, and newborn controls are significantly more active than adult controls. **(B)** Adults treated with MTD show significant naloxone-induced withdrawal, but newborns do not. **(A, B)** MTD dose is 10 mg/kg; *$p < 0.05$; **$p < 0.01$, by t-test (with Bonferroni correction in A for multiple testing).

juvenile mice, which have essentially no CNS barrier. Having established solid evidence of MTD effects on neonatal behaviors we set out to test this.

Effect of 6BN on Maternal Weight Gain During Pregnancy, and Pup Birthweight

For these studies most of the animals were tested with 5 mg/kg MTD and varying doses of 6BN. 5 mg/kg is at the midpoint of the dose-response curve (**Figure 3**). However, we also tested one cohort of animals at 7 mg/kg MTD and pooled the data with those at 5 mg/kg MTD (see Methods). First, we examined effects on pregnancy and birthweight. As shown in **Table 6**, MTD alone ("0 6BN") yielded a trend for decreased maternal weight gain, consistent with the MTD effect described in **Table 4**, but 6BN did not significantly reverse this effect. However, 6BN alone at 0.3 mg/kg (no MTD; n = 2) significantly increased maternal weight gain even above the saline controls. There were no effects on litter size or pup survival. As shown in **Table 7**, MTD had no effect on pup birthweight consistent with data in **Table 5**. However, 6BN in conjunction with MTD (even at the lowest 6BN dose of 0.025 mg/kg), or 6BN alone (at 0.3 mg/kg), caused a significant ~15% increase in pup birthweight above saline controls (F (4,149) = 5.48, $p < 0.001$, for overall effect of 6BN compared to saline controls, by ANOVA). In summary, this indicates an effect of 6BN to increase pup birthweight, and possibly maternal weight gain, independent of MTD treatment.

Effect of 6BN on Neonatal Withdrawal Behaviors

Next we tested whether 6BN could prevent neonatal withdrawal, at the doses of 5 & 7 mg/kg MTD as described above, applying the same four behavioral measures as used for the MTD dose-response. As shown in **Figure 6**, prenatal 6BN

dosing combined with MTD had a significant overall effect on three of the four measures, reducing the effect of MTD to baseline at the highest 6BN dose used (0.3 mg/kg). One measure, total number of calls, did not show any significant effect. Using the composite outcome method described above, we observed a significant overall effect of 6BN on reducing MTD-induced spontaneous withdrawal (**Figure 7A**). However, the composite score did not increase statistical sensitivity as observed in the MTD dose-response study (**Figure 3**). This is likely due to the number of calls. To test this, we removed that measure, reanalyzed the composite score data, and observed an increased statistical sensitivity based on detection of a significant effect at a lower dose of 6BN (by individual comparisons) than that observed for any single measure alone (compare asterisks in **Figures 6** and **7B**). These observations suggest that the two measures, number of calls and SNR, while increased by prenatal MTD exposure (**Figure 2**), may be differentially affected by 6BN with number of calls requiring higher doses for its suppression. We note here that for **Figure 7** the results were significant, as indicated, whether calculated based on all animals in each dosage group or based on litter averages, supporting that the results are not biased by a litter effect.

When composite outcome data shown in **Figure 7A** were fitted with an emax model (Bretz et al., 2005), the following parameters were obtained: e0 (i.e., the behavioral score at 6BN dose = 0) = 1.73 ± 0.70; emax (i.e, behavioral score at a maximal (infinite) 6BN dose) = - 2.724 ± 1.29; and ID50 = 0.020 ± 0.041. For the data shown in **Figure 7B** (i.e., a composite behavioral score without numcalls10), these parameters were estimated as follows: e0 = 1.59 ± 0.54; emax = -2.35 ± 0.91; and ID50 = 0.011 ± 0.023. The range in ID50 values for 6BN indicated by this analysis is similar to what we reported in morphine-dependent juvenile mice

TABLE 6 | The effect of 6BN on maternal weight gain and litter size.

6BN dose (mg/kg)	MTD dose (mg/kg)	n =	Start weight (g)	End weight (g)	Weight gain (g)	% Weight gain	AVG litter size	Total pup #	Total pup deaths
0	5,7	10 (8 at 5 MTD; 2 at 7 MTD)	1,190 ± 37	1,309 ± 53	118 ± 23**	9.7 ± 1.8	4.5 ± 0.4	48	1
0.025	5	6	1,252 ± 69	1,389 ± 61	137.5 ± 69	11.9 ± 5.1	4.2 ± 1.0	25	4
0.075	5,7	8 (6 at 5 MTD; 2 at 7 MTD)	1,230 ± 52	1,355 ± 51	125.9 ± 26	10.7 ± 2.5	4.3 ± 0.6	34	4
0.3	5,7	7 (5 at 5 MID; 2 at 7 MTD)	1,188 ± 35	1,349 ± 49	163 ± 24	13.7 ± 2.0	4.5 ± 0.7	27	1
0.3	0	2	1,134 ± 17	1,481 ± 25	347 ± 8*	30.6 ± 0.2	5.5 ± 0.5	11	0
0 (saline ctl)	0	5	1,188 ± 36	1,395 ± 31	206 ± 34	17.7 ± 3.4	5.8 ± 0.6	29	3

All values ± SEM. No overall significant effect of 6BN on maternal weight gain, by ANOVA *p < 0.05, compared to saline control by t-test; **p= 0.0664, compared to saline control, by t-test.

TABLE 7 | The effect of 6BN on pup birthweight.

6BN dose (mg/kg)	MTD dose (mg/kg)	Number of pups (n)	Number of litters	Birthweight (g)
0	5,7	49 (41 at 5 MTD; 8 at 7 MTD)	9	74.9 ± 1.6
0.025	5	21	5	85.8 ± 4.8
0.075	5,7	30 (22 at 5 MTD; 8 at 7 MTD)	8	84.2 ± 2.6
0.3	5,7	26 (17 at 5 MTD; 9 at 7 MTD)	6	89.8 ± 3.6[a]
0.3	0	11	2	88.5 ± 3.0[b]
0 (saline ctl)	0	28	6	75.6 ± 3.1

$F(4,149) = 5.484$, p = 0.00038 for overall effect of 6BN compared to saline controls, by ANOVA.
[a]p < 0.01 compared to saline ctl, by Dunnetts posthoc test; [b]p <0.01 compared to saline ctl, by t-test.

(0.02–0.04 mg/kg; (Oberdick et al., 2016)). However, more dosing data at the lower and higher ends will be needed to calculate a more accurate ID50 by fitting a 4-parametric log curve, which awaits further studies. Nevertheless, it is quite clear that 6BN is as effective in pregnant guinea pigs to prevent neonatal opioid dependence as it is in juvenile mice, against two different agonists with distinctly different PK properties. This is in spite of slow placental transit of 6BN and its consequent low fetal brain levels in guinea pigs.

MTD and 6BN Effects on Plasma Cortisol Levels

Previous studies have shown involvement of the hypothalamic-pituitary-adrenal (HPA) axis and neural-immune effects of maternal separation stress (MSS) (Hennessy 2003; Maken et al., 2010; Hennessy et al., 2011). Therefore, we tested the effect of MTD and 6BN on stress levels in newborns by measuring plasma cortisol. Plasma samples were collected from newborns 48 h after birth that had been exposed to prenatal MTD with and without 6BN, as well as 1 h of MSS just prior to sample collection. As shown in **Table 8** animals exposed to prenatal MTD had 46% greater cortisol levels compared to saline injected controls ($p <$ 0.05 by t-test). In addition, there was an overall significant effect of 6BN to prevent the MTD-induced increase in plasma cortisol (F (2,12) = 9.34; p = 0.0036 by ANOVA).

MTD and 6BN Effects in Adult Guinea Pigs

Previous studies have reported low potency of 6BN in adult rodents and monkeys for blocking opioid antinociception and for inducing withdrawal in opioid-dependent animals, with an ID50

ranging from 1 to 10 mg/kg (Wang et al., 2001; Wang et al., 2004; Raehal et al., 2005; Porter et al., 2002; Yancey-Wrona et al., 2009; Ko et al., 2006). Therefore, the finding that 6BN prevents withdrawal at much lower doses in juvenile mice (Oberdick et al., 2016) or in newborn guinea pigs exposed to a prenatal opioid (this study) was unexpected. Here we tested whether 6BN could prevent withdrawal in adult guinea pigs, which, as described above in PK studies, show relatively poor brain entry of 6BN (**Figure 1**). As adult non-pregnant guinea pigs tolerate MTD better than the pregnant animals, we tested the ability of 6BN to prevent naloxone-induced withdrawal after 3 days of exposure to MTD at 10 mg/kg, twice the dose of MTD we used for the 6BN study in pregnant animals. In fact, when co-administered with MTD, 6BN reduced withdrawal by 71% at the dose of 0.03 mg/kg, the lowest dose showing a significant effect (**Figure 8A**). Using a dose-response emax model, the parameters for the exponential fitted curve are e0 = 19.6 ± 3.62; emax = 4.5 ± 5.3; and ED50 = 0.010 ± 0.015. The mobility fitted for animals treated with the maximal dose (emax) was compared to the saline control (2.2 ± 1.2) and cannot be distinguished from it (p = 0.86 by t-test), and thus 6BN effectively reduces the effect of MTD to levels of saline controls. The AIC (Akaike's 'An Information Criterion', (Akaike, 1974)) for this model is 166.77.

It should be noted that eight of the 26 animals (31%) used in the adult study showed at least one bout of sleep-related hypotonia after naloxone treatment, with an overall average of one bout per animal for the adult dataset (see **Supplementary Movie S3**). Thus, this behavior is much less prominent than for newborns (see above). However, while five of these animals had only cursory bouts, spending less than 5% of their time in the
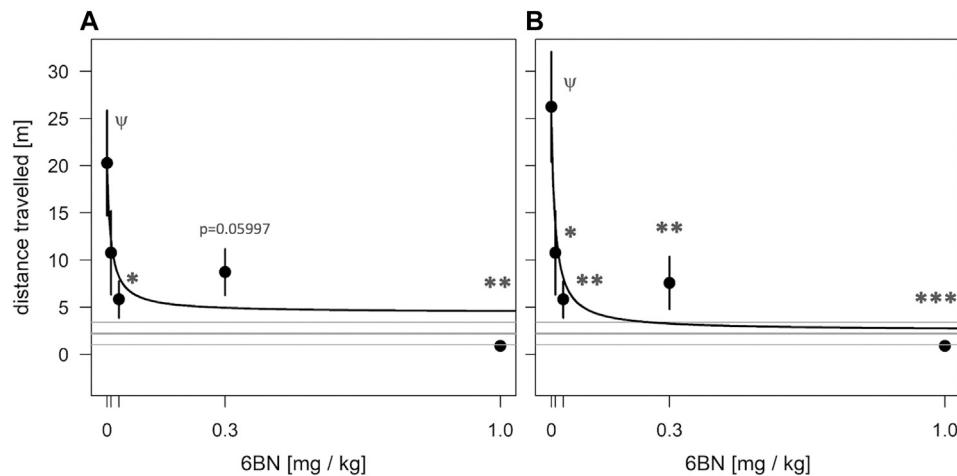
FIGURE 8 | 6BN has high potency to prevent opioid withdrawal in adult guinea pigs. Animals were treated for 3 days with one injection per day of 10 mg/kg MTD with varying doses of 6BN (0, 0.01, 0.03, 0.3, and 1 mg/kg). On day 4 animals were injected with naloxone and tested for 30 min in an open-field arena. **(A)** 6BN delivered at low doses with MTD prevents naloxone-induced withdrawal in adult animals. $p < 0.05$ for overall effect of 6BN, by linear trend test. **(B)** Same as A, but three animals with >15% of total arena-time in hypotonic posture were removed from dataset. $p < 0.01$ for overall effect of 6BN, by linear trend test. **(A)**, **(B)** *$p < 0.05$, **$p < 0.01$, ***$p < 0.001$, compared to "0 6BN" control, by Dunnett post-hoc test; ψ$p < 0.05$ compared to saline control (gray lines, mean ± SEM), by t-test.

arena in the prone hypotonic position, three of the animals spent greater than 15% of their time in this position. In addition, these three animals were either the highest or lowest in their respective groups for the measure of locomotion distance, suggesting an outsize influence on the data. To determine their effect on the overall curve shape and statistics we removed them from the dataset. As shown in **Figure 8B**, this allowed detection of a significant effect at all doses of 6BN, even at the lowest dose, 0.01 mg/kg. For this subset, the parameters for the exponential fit are e0 = 24.9 ± 3.7; emax = 2.54 ± 5.10; and ED50 = 0.010 ± 0.009 (with upper limit of 0.036 mg/kg with 99% confidence), and again, the mobility fitted for the maximal dose (emax) cannot be distinguished from saline controls ($p = 0.97$ by t-test). The AIC for this model is 139.23. The reduced AIC (better exponential fit) in this subset compared to the complete dataset, and the increased sensitivity of detecting a significant effect at a lower dose in this subset, suggests that hypotonia is linked to increased data variability in the naloxone dataset. We did not observe a similar improvement in detection of 6BN effects in the newborn naloxone dataset using this approach (not shown),

which we attribute to the much greater pervasiveness of this behavior in newborns.

DISCUSSION

In this study we can draw two major conclusions. First, we have shown that prenatal MTD exposure aggravates classic maternal separation behaviors in newborn guinea pigs, including locomotor and vocal behaviors. It also significantly increases plasma cortisol in newborns, an indicator of an activated brain HPA axis. These results suggest a likely conduit for prenatal opioids and opioid cessation at birth to affect later-life brain development and behavior, through HPA activation, as shown in many studies of early life stress in diverse species from rodents to non-human primates (Suomi 1997; Murgatroyd et al., 2009; Delpech et al., 2016; Roque et al., 2016; Wang et al., 2020). It is also consistent with the recent suggestion that salivary cortisol in NOWS babies may be a suitable indicator of withdrawal severity (Rodriguez et al., 2019). In addition, animal studies, primarily in guinea pigs and monkeys, but also in rats, have suggested a role for endogenous opioids in the natural process of maternal attachment (Herman and Panksepp, 1978; Kehoe and Blass, 1986; Kalin et al., 1988). This suggests that cessation of an opioid at birth after chronic exposure *in utero* may result in an increased drive or craving for the opioid receptor stimulation provided by maternal contact; hence, increased calling and searching. This may explain the effectiveness of parent or surrogate rooming-in, breast feeding, and kangaroo care therapies for reducing ICU stay times and reducing the need for postnatal opioids in the clinic (Dryden et al., 2009; Maguire, 2014; Wachman et al., 2018).

The second major finding is that 6BN, when delivered together with the agonist, can prevent dependence-related behaviors in

TABLE 8 | 6BN prevents the increase in cortisol by prenatal MTD.

6BN treatment	Cortisol (ng/ml)
0	1,399
0.075	1,094
0.3	859[a]
Saline ctl	958[b]

All pups treated with prenatal MTD (5 or 7 mg/kg) for 15 days with 6BN at the indicated dose, or with saline (no MTD, no 6BN); n = 5 pups from 3 litters per group = 3 pups at 5 mg/kg MTD and two pups at 7 mg/kg MTD in each group. F(2,12) = 9.34, p = 0.0036 for overall effect of 6BN, by ANOVA.
[b]p<0.05, by t-test.
[a]p<0.01 by Dunnett's posthoc analysis, all comparisons to 0 6BN condition.

newborns exposed to extended periods of prenatal MTD - at 6BN doses unlikely to induce maternal or fetal withdrawal, or to interfere with opioid analgesia or use management. For example, the ID50 of 6BN for inducing withdrawal in opioid dependent rodents is in the range 1–10 mg/kg (Wang et al., 2001; Wang et al., 2004), or 1, 1.3 or 2.4 mg/kg (Porter et al., 2002; Raehal et al., 2005; Yancey-Wrona et al., 2009) for inhibition of opioid antinoception in rodents, or in the range of 0.3–1 mg/kg for interference of antinociception and induction of withdrawal in rhesus monkeys (Ko et al., 2006). In addition, the potency of prenatally delivered 6BN to prevent withdrawal in newborn guinea pigs, with an estimated ID50 in the range of 0.01–0.02 mg/kg, is in a similar range as we observed in juvenile mice treated with morphine (Oberdick et al., 2016). This high potency is in spite of the fact that MTD has a much longer half-life than 6BN, 12 h for MTD in guinea pigs (Pak and Ecobichon, 1978) vs. ~2 h for 6BN (this study), and that 6BN has slow placental transit in guinea pigs (this study). Consistent with this result, we find that 6BN also has high potency to prevent withdrawal in adult guinea pigs, in which 6BN is relatively excluded from the CNS (also this study). A very similar observation has recently been made in adult mice using morphine rather than methadone as an agonist (Z. Jim Wang, U. Illinois–Chicago; personal communication).

From this evidence we conclude that barrier mechanisms at the placenta or BBB do not impede the potency of 6BN in preventing neonatal or adult dependence, and other mechanisms may be in play to explain its relatively low potency to induce withdrawal in dependent animals or to interfere in opioid analgesia. One possibility is a greater role of the peripheral nervous system in driving withdrawal behaviors than is currently appreciated. For example, recent studies have shown that auricular stimulation of cranial nerves reduces withdrawal symptoms, and may act by reducing sympathetic activity ("fight or flight") that is increased during withdrawal, favoring parasympathetic predominance and reducing physical withdrawal symptoms (Qureshi et al., 2020). Preferential blockade of opioid actions on cranial nerves by 6BN could maintain normal function of these peripheral neurons so that when the opioid ceases there is reduced sympathetic imbalance. Alternatively, 6BN may interact in novel ways with the opioid receptor, possibly binding with high affinity to a distinct receptor conformation involved in the development of dependence. For example, Jeske (Jeske 2019) has proposed a receptor model with distinct peripheral and central mu opioid receptor forms that could in part account for the observed peripheral selectivity of 6BN, even in rhesus monkeys where it readily penetrates the adult's brain (J. Oberdick, unpublished results; see also (Sadee et al., 2020)). It appears that our current understanding of the molecular pharmacology of opioid receptors is incomplete–requiring a new approach to account for the high potency of 6BN in selectively preventing opioid withdrawal behavior in dependent animals, as observed here and in mice (Oberdick et al., 2016; Sadee et al., 2020).

Several novel features of the guinea pig model are worthy of discussion. First, the behavior of guinea pig newborns enables a focus on maternal separation stress immediately after birth. This two-stage process mimics what is observed in non-human primates (Hennessy 2003), and since it clearly involves the HPA axis, neuroinflammatory mechanisms are immediately suggested (Maken et al., 2010; Hennessy et al., 2011). In contrast, rats at birth show only modest behaviors, which, while statistically significant, are not immediately relatable to specific CNS neuronal pathways (Robinson and Wallace, 2001; Griffin et al., 2019). Not until well after birth, in the period PD7–PD10, do rat pups show evidence of a clear maternal separation stress response. As in guinea pig newborns this response in PD7-10 rats appears to be dependent on endogenous opioids (Kehoe and Blass, 1986), and it is aggravated by daily opioid treatments for the first postnatal week with cessation on PD7 (Barr and Wang, 1992). While these 7-day old rat pups may serve as a reasonable model for some features of NOWS, they lack the temporal feature of withdrawal that is initiated due to opioid cessation at birth.

The second novel feature in newborn guinea pigs is the close apposition of two behaviors, locomotion and vocalization, that are at the heart of the maternal separation stress response. These two behaviors are developmentally coordinated in a manner linked to arousal, and thus their coordination is highly influenced by environment (Gustison et al., 2019). Thus, multiple but distinct measures of each of these two behaviors may be useful tools for dissecting neuronal pathways. In particular, SNR (or "loudness") may be a vocalization feature that most closely relates to animal stress, and the magnitude of increase in stress vocalization that we observed when comparing saline controls to prenatal opioid exposed newborns, ~4 db, is equivalent to the magnitude of change observed during castration of unanesthetized newborn pigs in a farm setting (von Borell et al., 2009). This comparison may highlight that newborns exposed to prenatal opioids are under considerable stress. Also, while number of calls is a measure most often used to assess changes in vocalization in mouse reverse genetics studies (e.g., (Jamain et al., 2008)), the fact that this measure is not significantly suppressed by 6BN in newborn guinea pigs exposed to prenatal opioids, while plasma cortisol levels are suppressed, may suggest that counts of number of calls are not as relevant to stress as is SNR.

A third novel feature of the guinea pig as observed here is the sleep-related hypotonia induced by naloxone. The behavior is often accompanied by yawning, which supports that it is sleep-related. The observation that it is much more prominent in newborns than adults may be due to differences in physiology, but the fact that it is observed even in pups that received prenatal saline instead of MTD may suggest a strong influence of endogenous opioids, which as described above likely drives natural infant-maternal attachment (Herman and Panksepp, 1978; Kehoe and Blass, 1986; Kalin et al., 1988). In adults it appears that the relatively less prevalent version of this naloxone-induced behavior is specific to animals dependent on exogenous opioids, but more studies are needed. Disrupted sleep patterns is a well-described feature of NOWS in the clinic (O'Brien and Jeffery, 2002; Wachman et al., 2020), and therefore circadian activity studies may be of value in the future to study the effects on sleep and wakefulness in guinea pig newborns exposed to prenatal opioids. In addition, the naloxone-induced hypotonia behavior of newborns appears remarkably similar to the passive/depressive behaviors characteristic of older juvenile pre-weaning guinea pig pups upon maternal separation, which may be indicative of a broader separation-induced sickness-like behavior

with neuroinflammatory origins as previously described (Hennessy et al., 2011; Hennessy et al., 2015). In future studies one could explore whether this behavior might be detected in newborns without naloxone after more extended periods of separation, and if so, whether this effect, and its sensitization by repeated bouts of maternal separation (Hennessy et al., 2011; Hennessy et al., 2015), is aggravated by prenatal opioids.

The final feature worthy of discussion is the ability to combine these behaviors into a single composite score, at the level of each individual animal. This feature may allow for both increased efficiency of animal usage in terms of statistical detection, but also, by providing a more robust read-out of withdrawal, it may allow better understanding of the relationship between neonatal withdrawal severity and developmental consequences in later-life.

In conclusion, here we report a robust behavioral model of NOWS in pregnant guinea pigs, and we take a first step toward testing 6BN as a preventive therapy for NOWS, with a focus on newborn outcomes. Further studies are needed to determine the limits of 6BN potency, such as at higher doses of MTD, or with other commonly used opioids in the clinic such as buprenorphine to treat OUD, or morphine or oxymorphone for pain. In addition, further studies are needed to specifically examine, in adult and pregnant guinea pigs, the levels of 6BN needed to either induce withdrawal in opioid dependent animals or to interfere with opioid antinociception. 6BN has been universally shown to have low potency for these actions in both adult rodents and monkeys, and it will be important to contrast this with low levels needed to prevent neonatal withdrawal in a single animal model. In addition, it remains to be seen whether initiation of MTD and 6BN combined therapy in pregnant animals that are already MTD-dependent, a condition likely to apply in the clinic, causes any untoward maternal or fetal effects related to withdrawal. Thus, in future studies we will focus much more on effects of opioids and 6BN on maternal outcomes.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

ETHICS STATEMENT

The animal study was reviewed and approved by The Ohio State University IACUC.

AUTHOR CONTRIBUTIONS

Performing experiments: JO, AS, SA, and VM guinea pig husbandry, tissue collection, and behavior; KH and KD mass spectrometry. Experimental design: JO, KB, WS, and MP Scoring of behavior by analysis of video recordings: AL (vocalization) and JF (locomotion). Statistical analysis: MC and MP (pharmacokinetics) and KS (behavior). Manuscript writing and revision, and financial support: JO, SA, AS, KS, AL, KB, KO, MP, and WS.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2020.613328/full#supplementary-material.

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Conflict of Interest: WS is Chief Scientific Officer of Aether Therapeutics and holds share in Aether.